

This article was downloaded by:

On: 23 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713617200>

Synthetic Studies on Sialoglycoconjugates 52: Synthesis of Sialyl Lewis X Analogs Containing Azidoalkyl Groups at the Reducing End

Akira Hasegawa^a; Koshiro Fushimi^a; Hideharu Ishida^a; Makoto Kiso^a

^a Department of Applied Bioorganic Chemistry, Gifu University, Gifu, Japan

To cite this Article Hasegawa, Akira , Fushimi, Koshiro , Ishida, Hideharu and Kiso, Makoto(1993) 'Synthetic Studies on Sialoglycoconjugates 52: Synthesis of Sialyl Lewis X Analogs Containing Azidoalkyl Groups at the Reducing End', *Journal of Carbohydrate Chemistry*, 12: 8, 1203 – 1216

To link to this Article: DOI: 10.1080/07328309308020128

URL: <http://dx.doi.org/10.1080/07328309308020128>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

**SYNTHETIC STUDIES ON SIALOGLYCOCONJUGATES 52:
SYNTHESIS OF SIALYL LEWIS X ANALOGS CONTAINING
AZIDOALKYL GROUPS AT THE REDUCING END**

Akira Hasegawa, Koshiro Fushimi, Hideharu Ishida and Makoto Kiso

Department of Applied Bioorganic Chemistry, Gifu University,
Gifu 501-11, Japan

Received April 10, 1993 - Final Form July 21, 1993

ABSTRACT

Stereocontrolled synthesis of sialyl Le^x epitope analogs in which the terminal *N*-acetylglucosamine residue of sialyl Le^x determinant is replaced by a D-glucopyranose residue containing β-glycosidically linked azidoalkyl groups is described. Glycosylation of 2-(trimethylsilyl)ethyl *O*-(2,6-di-*O*-benzoyl-3,4-*O*-isopropylidene-β-D-galactopyranosyl)-(1→4)-2,6-di-*O*-benzoyl-β-D-glucopyranoside (**2**), prepared from 2-(trimethylsilyl)ethyl β-lactoside (**1**) by 3,4-*O*-isopropylideneation and selective-*O*-benzoylation, with methyl 2,3,4-tri-*O*-benzyl-1-thio-β-L-fucopyranoside (**3**) gave the desired α-glycoside **4**, which was converted by *O*-deisopropylideneation into **7**, and *via O*-debenzoylation, selective 2,6,6'-tri-*O*-benzoylation and *O*-deisopropylideneation into **8**, respectively. *N*-Iodosuccinimide (NIS)-TfOH-promoted glycosylation of **7** or **8** with methyl (phenyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-2-thio-D-glycero-D-galacto-2-nonulopyranosid)onate (**9**) afforded the desired tetrasaccharides **10** and **11**.

Compound **11** was converted into the α-trichloroacetimidate **14** *via* reductive removal of the benzyl groups, *O*-acetylation, removal of the 2-(trimethylsilyl)ethyl group and treatment with trichloroacetonitrile. Coupling of **14** with 2-azidoethanol, 8-azidooc-tanol, and 2-[2-(2-azidoethoxy)ethoxy]ethanol, gave the desired β-glycosides **15-17**, respectively. *O*-Deacylation of **12**, **15-17** and subsequent hydrolysis of the methyl ester group yielded the title compounds.

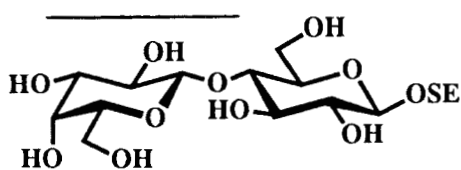
INTRODUCTION

Recently, it has been demonstrated¹⁻³ that the selectin family, such as E-selectin (endothelial leukocyte adhesion molecule-1, ELAM-1), P-selectin (granule membrane protein, GMP-140), and L-selectin (LECAM-1), recognizes the sialyl Le^x determinant, α-Neu5Ac-(2→3)-β-D-Gal-(1→4)-[α-L-Fuc-(1→3)]-β-GlcNAc, which is found as the terminal carbohydrate structure of both cell membrane glycolipids and glycoproteins.

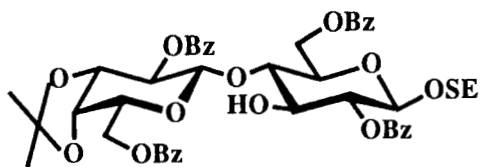
Previously, we have reported the synthesis of sialyl Le^x ganglioside (a hexasaccharide⁴ and pentasaccharide⁵), α -sialyl-(2 \rightarrow 6)-Le^x ganglioside,⁶ and the analogs,⁷ and have examined their recognition activity by selectin family. The data^{3,7b,8-11} showed that the fucose structure was required for full recognition, the side-chain structure of the sialic acid residue was not critical for the activity, and α -sialyl-(2 \rightarrow 6)Le^x ganglioside was not recognized at all. Recently, it has been shown that the terminal *N*-acetylglucosamine residue of sialyl Le^x oligosaccharide could be replaced¹² by D-glucopyranose without decreasing the activity, indicating the more detailed structural requirement necessary for selectin recognition. In view of these facts, we describe herein the stereocontrolled synthesis of sialyl Le^x analogs in which the terminal *N*-acetylglucosamine is replaced by D-glucopyranose residues containing β -glycosidically linked azidoalkyl groups. Availability of these analogs is directed toward elucidation of cell-cell adhesion mechanism and application of the sialyl Le^x epitope structure to medicinal use.

RESULTS AND DISCUSSION

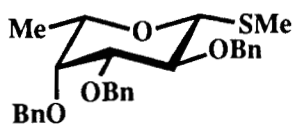
Treatment of 2-(trimethylsilyl)ethyl *O*- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-glucopyranoside¹³ (**1**) with 2,2-dimethoxypropane (2.0 equiv to **1**) in *N,N*-dimethylformamide (DMF) containing *p*-toluenesulfonic acid monohydrate for 3 h at 80 °C, afforded the 3,4-*O*-isopropylidene derivative, which, on selective 2,6,2',6'-*O*-benzoylation with benzoyl chloride, gave the desired glycosyl acceptor **2** in 54% yield; significant signals in the ¹H NMR spectrum were a one-proton doublet of doublets at δ 5.36 ($J_{1,2} = 8.1$ Hz, $J_{2,3} = 9.5$ Hz, H-2 for Gal), a triplet at δ 5.52 due to H-2 of Glc, and twenty aromatic protons due to four phenyl groups at δ 7.31-8.26, indicating the structure assigned. Dimethyl(methylthio)sulfonium triflate¹⁴ (DMTST)-promoted glycosylation of **2** with methyl 2,3,4-tri-*O*-benzyl-1-thio- β -L-fucopyranoside^{4b,15} (**3**) in benzene for 3 h at 0 °C gave the desired α -glycoside **4** in 90% yield. The ¹H NMR signal for the Fuc unit in **4** [δ 5.62 ($J_{1,2} = 4.1$ Hz, H-1)] indicated the glycosidic configuration to be α . Hydrolytic removal of the isopropylidene group in **4** under mild, acidic conditions gave the glycosyl acceptor **7** in 90% yield. Alternatively, the other glycosyl acceptor **8**, in which the C-2, C-3 and C-4 hydroxyl groups of the galactose residue are not protected, designed to give a better yield for the next α -glycosylation using a sialic acid glycosyl donor, was prepared in good yield *via O*-debenzoylation of **4** and selective *O*-benzoylation followed by hydrolytic removal of the isopropylidene group; significant signals in the ¹H NMR spectrum were two one-proton doublets of doublets (δ 4.12, $J_{1,2} = 7.7$ Hz, $J_{2,3} = 9.3$ Hz, H-2 for Gal; δ 5.18, $J_{1,2} = 8.1$ Hz, $J_{2,3} = 9.3$ Hz, H-2 for Glc) and thirty aromatic protons due to six phenyl groups, indicating the structure assigned.



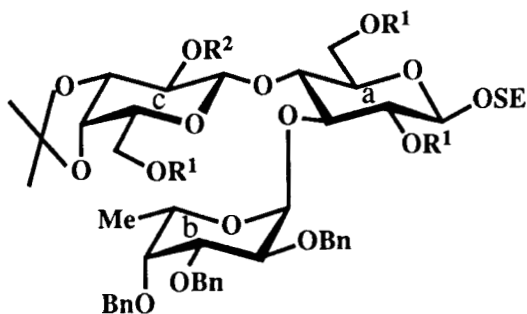
1



2

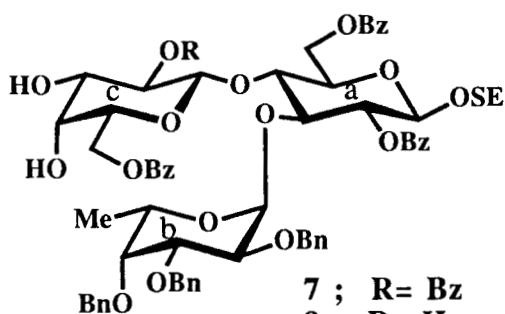


3

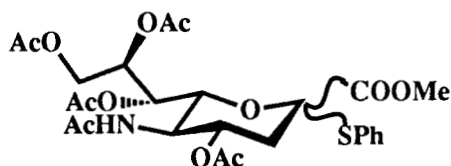


SE ; 2-(trimethylsilyl)ethyl
Bz ; benzoyl
Bn ; benzyl

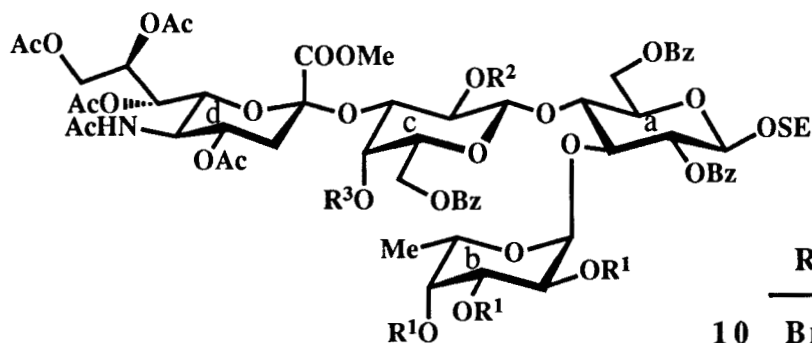
| | R ¹ | R ² |
|---|----------------|----------------|
| 4 | Bz | Bz |
| 5 | H | H |
| 6 | Bz | H |



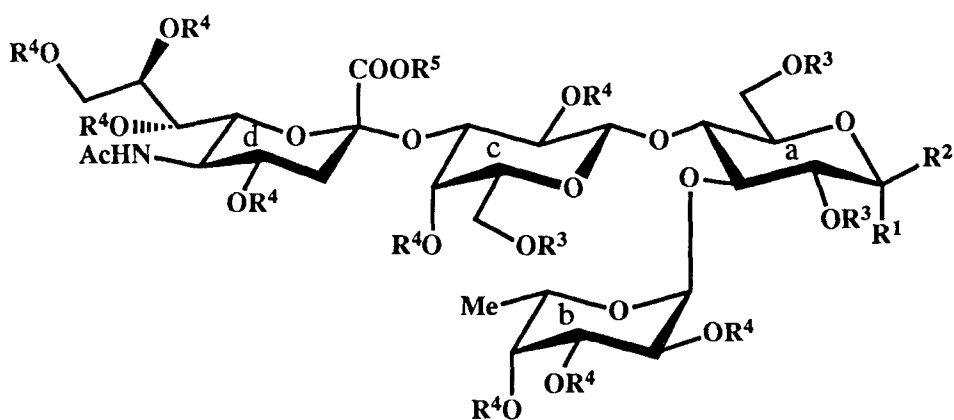
7 ; R = Bz
8 ; R = H



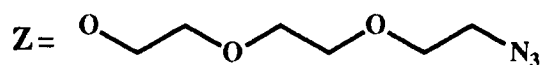
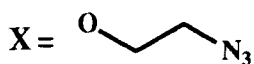
9



| | R ¹ | R ² | R ³ |
|----|----------------|----------------|----------------|
| 10 | Bn | Bz | H |
| 11 | Bn | H | H |
| 12 | Ac | Ac | Ac |



| | R ¹ | R ² | R ³ | R ⁴ | R ⁵ |
|----|-----------------------------|----------------|----------------|----------------|----------------|
| 13 | H, OH | | Bz | Ac | Me |
| 14 | OC(=NH)CCl ₃ , H | | Bz | Ac | Me |
| 15 | H | X | Bz | Ac | Me |
| 16 | H | Y | Bz | Ac | Me |
| 17 | H | Z | Bz | Ac | Me |
| 18 | H | X | H | H | H |
| 19 | H | Y | H | H | H |
| 20 | H | Z | H | H | H |
| 21 | H | OSE | H | H | H |



The glycosylation of **7** or **8** with methyl (phenyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-2-thio-D-glycero- α -D-galacto-2-nonulopyranosid)onate¹⁶ (**9**) in the presence of *N*-iodosuccinimide (NIS)-trifluoromethanesulfonic acid (TfOH) as the glycosyl promoter¹⁷ and powdered molecular sieves 3Å (MS-3Å) in CH₃CN overnight at -35 °C, afforded the corresponding desired α -glycosides **10** (41%) and **11** (58%), respectively. As expected, compound **8** having the three adjacent hydroxyl groups at C-2, -3, -4 of the galactose residue behaved as a suitable glycosyl acceptor for the selective α -glycosylation of Neu5Ac to the C-3, similar to what have been observed previously.^{17c,18} The structure of the glycosides **10** and **11** were unambiguously proved by 270 MHz ¹H NMR spectroscopy. The observed chemical constants of Neu5Ac unit in the glycosides for H-3_{eq} (δ 2.67, $J_{\text{gem}} = 12.8$ Hz, $J_{3\text{eq},4} = 4.6$ Hz, **10**; δ 2.54, $J_{\text{gem}} = 12.6$ Hz, $J_{3\text{eq},4} = 4.3$ Hz, **11**), H-4 (δ 4.95, **10**; δ 4.90, **11**), and H-7 (δ 5.43, $J_{6,7} = 2.0$ Hz, $J_{7,8} = 9.3$ Hz, **10**; δ 5.28, $J_{6,7} = 2.3$ Hz, $J_{7,8} = 9.5$ Hz, **11**) are characteristic of α -glycosidic linkages^{18b,c} of Neu5Ac. Other ¹H NMR data are given in Experimental Section and are consistent with the structures assigned. Catalytic hydrogenolysis (10% Pd-C) of the benzyl groups of **11** in ethanol-acetic acid for 48 h at 40 °C and subsequent *O*-acetylation gave the per-*O*-acyl compound **12** in 89% yield, which, on *O*-deacylation with sodium methoxide in methanol and subsequent saponification of the methyl ester group, yielded the desired sialyl Le^x epitope analog **21** in quantitative yield. Treatment^{4,13a} of **12** with trifluoroacetic acid in CH₂Cl₂ for 3 h at 0 °C gave the 1-hydroxy compound **13** in 95% yield. Compound **13** when treated with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) for 2 h at 0 °C gave the α -trichloroacetimidate **14** in 87% yield. The ¹H NMR data for Glc unit in **14** [δ 6.35 ($J_{1,2} = 3.8$ Hz, H-1), 8.58 (C=NH)] indicated the imidate to be α .

The glycosylation of 2-azidoethanol, 8-azidoethanol, or 2-[2-(2-azidoethoxy)ethoxy]ethanol, with **14** thus obtained, in CH₂Cl₂ in the presence of boron trifluoride etherate¹⁹ for 8 h at 0 °C afforded the expected β -glycosides **15-17** in high yields, respectively. The ¹H NMR data for the Glc unit of **15-17** [δ 4.78-4.84 ($J_{1,2} = 7.5\sim 7.8$ Hz, H-1)] indicated the glycosidic linkages to be β . Finally, *O*-deacylation of **15-17** with sodium methoxide in methanol, with subsequent saponification of the methyl ester group, yielded the corresponding, desired sialyl Le^x epitope analogs (**18-20**) in good yields, respectively.

EXPERIMENTAL

General Procedures. Specific rotations were determined with a Union PM-201 polarimeter at 25 °C, and IR spectra were recorded with a Jasco IRA-100 spec-

trophotometer. ^1H NMR spectra were recorded with a JEOL JNM-GX 270 spectrometer. Preparative chromatography was performed on silica gel (Wako Co., 200 mesh) with the solvent systems specified. Concentrations were conducted *in vacuo*.

2-(Trimethylsilyl)ethyl *O*-(2,6-Di-*O*-benzoyl-3,4-*O*-isopropylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,6-di-*O*-benzoyl- β -D-glucopyranoside (2). To a solution of 2-(trimethylsilyl)ethyl β -lactoside¹³ (1; 8.8 g, 20 mmol) in *N,N*-dimethylformamide (DMF, 50 mL) were added 2,2-dimethoxypropane (5.3 mL, 40 mmol) and *p*-toluenesulfonic acid monohydrate (100 mg), and the mixture was stirred for 3 h at 80 °C and neutralized with Amberlite IR-410 (HO⁻) resin. The resin was filtered off and washed with MeOH, and the combined filtrate and washings were concentrated. Column chromatography (50:1 CH₂Cl₂-MeOH) of the residue on silica gel (300 g) gave the 3',4'-*O*-isopropylidene derivative (6.1 g, 64%) as an amorphous mass, which was used for the next reaction. To a solution of the isopropylidene derivative (2.0 g, 4.1 mmol) in dry pyridine (6.7 mL) and CH₂Cl₂ (10 mL), cooled to -40 °C, was added dropwise, with stirring, a solution of benzoyl chloride (2.4 mL, 20.5 mmol) in CH₂Cl₂ (20 mL), and the stirring was continued for 6 h at -40 °C. Methanol (5 mL) was added to the mixture, concentrated and extracted with CH₂Cl₂. The extract was successively washed with 2M HCl and H₂O, dried (Na₂SO₄) and concentrated. Column chromatography (300:1 CH₂Cl₂-MeOH) of the residue on silica gel (50 g) gave **2** (2.0 g, 54%) as an amorphous mass: $[\alpha]_{\text{D}}^{25} +15.6^\circ$ (*c* 0.4, CHCl₃); IR (KBr) 3600-3300 (OH), 1730 and 1250 (ester), and 840 (TMS, Me₂C), and 710 cm⁻¹ (Ph); ^1H NMR (CDCl₃) δ 0.98 (m, 2H, Me₃SiCH₂CH₂), 1.51, 1.79 (2s, 6H, Me₂C), 3.65, 4.02 (2m, 2H, Me₃SiCH₂CH₂), 4.14 (broad t, 1H, J_{2,3} = J_{3,4} = 8.2 Hz, H-3), 4.71 (d, 1H, J_{1,2} = 8.1 Hz, H-1'), 4.82 (d, 1H, J_{1,2} = 8.2 Hz, H-1), 5.36 (dd, 1H, J_{2,3} = 9.5 Hz, H-2'), 5.52 (broad t, 1H, H-2), and 7.31-8.26 (m, 20H, 4Ph).

Anal. Calcd for C₄₈H₅₄O₁₅Si (899.0): C, 64.13; H, 6.05. Found: C, 63.86; H, 5.95.

2-(Trimethylsilyl)ethyl *O*-(2,6-Di-*O*-benzoyl-3,4-*O*-isopropylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-*O*-benzoyl- β -D-glucopyranoside (4). To a solution of **2** (1.06 g, 1.2 mmol) and methyl 2,3,4-tri-*O*-benzyl-1-thio- β -L-fucopyranoside^{4b, 15} (**3**; 818 mg, 1.8 mmol) in dry benzene (5 mL) were added powdered molecular sieves 4Å (MS-4Å, 3 g), and the mixture was stirred for 5 h at room temperature, then cooled to 0 °C. Dimethyl(methylthio)sulfonium triflate¹⁴ (DMTST; 1.5 g, 5.3 mmol) was added to the stirred mixture, and the stirring was continued for 3 h at 7 °C; the course of the reaction was monitored by TLC. Methanol (2 mL) and triethylamine (1 mL) were added to the mixture, concentrated, and extracted with CH₂Cl₂. The extract was washed with H₂O,

dried (Na_2SO_4) and concentrated. Column chromatography (400:1 CH_2Cl_2 -MeOH) of the residue on silica gel (80 g) gave **4** (1.4 g, 90%) as an amorphous mass: $[\alpha]_{\text{D}} -25.0^\circ$ (c 0.16, CHCl_3); ^1H NMR (CDCl_3) δ 0.90 (m, 2H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.46, 1.69 (2s, 6H, Me_2C), 1.52 (d, 3H, $J_{5,6} = 6.6$ Hz, H-6b), 3.56, 4.32 (2m, 2H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 4.95 (d, 1H, $J_{1,2} = 6.1$ Hz, H-1a), 5.44 (broad t, 1H, $J_{1,2} = J_{2,3} = 8.2$ Hz, H-2c), 5.61 (broad t, 1H, $J_{1,2} = J_{2,3} = 8.8$ Hz, H-2a), 5.63 (d, 1H, $J_{1,2} = 3.6$ Hz, H-1b), and 7.13-8.38 (m, 35H, 7Ph).

Anal. Calcd for $\text{C}_{75}\text{H}_{82}\text{O}_{19}\text{Si}$ (1315.6): C, 68.48; H, 6.28. Found: C, 68.46; H, 6.37.

2-(Trimethylsilyl)ethyl O-(3,4-O-Isopropylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-[O-(2,3,4-tri-O-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-D-glucopyranoside (5). To a solution of **4** (2.27 g, 1.73 mmol) in MeOH (20 mL) and CH_2Cl_2 (3 mL) was added sodium methoxide (100 mg) and the mixture was kept at 60°C ; the course of the reaction was monitored by TLC. After completion of the reaction, the mixture was neutralized by Amberlite IR-120 (H^+) resin. The solution was concentrated. Column chromatography (30:1 CH_2Cl_2 -MeOH) of the residue on silica gel afforded **5** (1.34 g, 86%) as an amorphous mass: $[\alpha]_{\text{D}} -71.5^\circ$ (c 0.2, MeOH); ^1H NMR (CD_3OD) δ 0.98 (m, 2H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.04, (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.27, 1.33 (2s, 6H, Me_2C), 3.44 (dd, 1H, $J_{1,2} = 7.9$ Hz, $J_{2,3} = 9.0$ Hz, H-2c), 4.27 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1a), 5.67 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1b), and 7.21-7.93 (m, 15H, 3Ph).

Anal. Calcd for $\text{C}_{47}\text{H}_{66}\text{O}_{15}\text{Si}$ (899.1): C, 62.79; H, 7.40. Found: C, 62.65; H, 7.57.

2-(Trimethylsilyl)ethyl O-(6-O-Benzoyl-3,4-O-isopropylidene- β -D-galactopyranosyl)-(1 \rightarrow 4)-[O-(2, 3, 6-tri-O-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-O-benzoyl- β -D-glucopyranoside (6). To a solution of **5** (621 mg, 0.69 mmol) in pyridine (1 mL) and CH_2Cl_2 (10 mL), cooled to -40°C , was added a solution of benzoyl chloride (0.48 mL, 4.1 mmol), and the mixture was stirred for 6 h at -40°C and then worked-up, as described for **2**, to give **6** (620 mg, 74%) as an amorphous mass: $[\alpha]_{\text{D}} -41.5^\circ$ (c 0.2, CHCl_3); ^1H NMR (CDCl_3) δ 0.98 (m, 2H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.35 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.60, 1.64 (2s, 6H, Me_2C), 3.74 (t, 1H, $J_{1,2} = J_{2,3} = 9.4$ Hz, H-2c), 5.29 (dd, 1H, $J_{1,2} = 8.4$ Hz, $J_{2,3} = 7.3$ Hz, H-2a), 5.38 (d, 1H, $J_{1,2} = 3.1$ Hz, H-1b), and 7.24-8.21 (m, 30H, 6Ph).

Anal. Calcd for $\text{C}_{68}\text{H}_{78}\text{O}_{18}\text{Si}$ (1211.4): C, 67.42; H, 6.49. Found: C, 67.39; H, 6.29.

2-(Trimethylsilyl)ethyl O-(2,6-Di-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[O-(2, 3, 4-tri-O-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-O-ben-

zoyl- β -D-glucopyranoside (7). A solution of **4** (240 mg, 0.18 mmol) in aqueous 80% AcOH was heated overnight at 50 °C and concentrated. Column chromatography (200:1 CH₂Cl₂-MeOH) of the residue on silica gel gave **7** (209 mg, 90%) as an amorphous mass: [α]_D -37.5° (c 0.34, CHCl₃); ¹H NMR (CDCl₃) δ 0.98 (m, 2H, Me₃SiCH₂CH₂), 1.47 (d, 3H, J_{5,6} = 6.0 Hz, H-6b), 5.45 (dd, 1H, J_{1,2} = 7.9 Hz, J_{2,3} = 8.2 Hz, H-2c), 5.57 (d, 1H, J_{1,2} = 3.6 Hz, H-1b), 5.63 (t, 1H, J_{1,2} = J_{2,3} = 8.4 Hz, H-2a), and 7.13-8.38 (m, 35H, 7Ph).

Anal. Calcd for C₇₂H₇₈O₁₉Si (1275.5): C, 67.80; H, 6.16. Found: C, 67.54; H, 6.32.

2-(Trimethylsilyl)ethyl O-(6-O-Benzoyl- β -D-galactopyranosyl)-[O-(2,3,4-tri-O-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-O-benzoyl- β -D-glucopyranoside (8). A solution of **6** (400 mg, 0.33 mmol) in aqueous 80% AcOH (40 mL) was stirred overnight at room temperature and concentrated. Column chromatography (80:1 CH₂Cl₂-MeOH) of the residue on silica gel (40 g) gave **8** (351 mg, 91%) as an amorphous mass: [α]_D -51.5° (c 0.2, CHCl₃); IR (KBr) 3450 (OH), 1730 and 1230 (ester), and 860 and 840 (TMS), and 750 and 710 cm⁻¹ (Ph); ¹H NMR (CDCl₃) δ 0.98 (m, 2H, Me₃SiCH₂CH₂), 1.27 (d, 3H, J_{5,6} = 6.4 Hz, H-6b), 3.67 (t, 1H, J_{2,3} = J_{3,4} = 9.3 Hz, H-3a), 3.68 (dd, 1H, J_{2,3} = 9.3 Hz, J_{3,4} = 4.5 Hz, H-3c), 3.96 (t, 1H, J_{4,5} = 9.5 Hz, H-4a), 4.11 (dd, 1H, J_{1,2} = 2.9 Hz, J_{2,3} = 12.5 Hz, H-2b), 4.12 (dd, 1H, J_{1,2} = 7.7 Hz, H-2c), 4.27 (d, 1H, H-1c), 4.59 (d, 1H, J_{1,2} = 8.1 Hz, H-1a), 5.18 (dd, 1H, J_{2,3} = 9.3 Hz, H-2a), 5.34 (d, 1H, H-1b), and 7.16-8.08 (m, 30H, 6Ph).

Anal. Calcd for C₆₅H₇₄O₁₈Si (1171.4): C, 66.65; H, 6.37. Found: C, 66.40; H, 6.33.

2-(Trimethylsilyl)ethyl O-(Methyl 5-Acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2 \rightarrow 3)-O-(2,6-di-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[O-(2,3,4-tri-O-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-O-benzoyl- β -D-glucopyranoside (10). To a solution of **7** (120 mg, 94 μ mol) and **9** (115 mg, 197 μ mol) in dry CH₃CN (2.5 mL) were added molecular sieves 3Å (MS-3Å, 1.5 g) and the mixture was stirred for 5 h at room temperature and cooled to -40 °C. *N*-Iodosuccinimide (120 mg, 0.54 mmol) and TfOH (12 μ L, 0.14 μ mol) were added to the reaction mixture under N₂ atmosphere and it was stirred overnight at -35 °C, neutralized with triethylamine, concentrated, and then extracted with CH₂Cl₂. The extract was successively washed with M Na₂S₂O₃ and water, dried (Na₂SO₄) and concentrated. Column chromatography (50:1 toluene-MeOH) of the residue on silica gel (40 g) gave **10** (67 mg, 41%) as an amorphous mass: [α]_D -14.0° (c 0.9, CHCl₃); IR (KBr) 3370 (OH, NH), 1740 and 1220 (ester), 1660 and

1540 (amide), 860 and 840 (TMS), and 720 cm^{-1} (Ph); ^1H NMR (CDCl_3) δ 0.98 (m, 2H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.56 (d, 3H, $J_{5,6} = 6.6$ Hz, H-6b), 1.71, 1.97, 2.02, 2.14, 2.21 (5s, 15H, AcN, 4AcO), 2.67 (dd, 1H, $J_{\text{gem}} = 12.8$ Hz, $J_{3\text{eq},4} = 4.6$ Hz, H-3 deq), 4.95 (m, 1H, H-4d), 5.43 (dd, 1H, $J_{6,7} = 2.0$ Hz, $J_{7,8} = 9.3$ Hz, H-7d), 5.55 (d, 1H, $J_{1,2} = 3.0$ Hz, H-1b), 5.72 (m, 1H, H-8d), and 7.13–8.36 (m, 35H, 7Ph).

Anal. Calcd for $\text{C}_{92}\text{H}_{105}\text{NO}_3\text{Si}$ (1748.9): C, 63.18; H, 6.05; N, 0.80. Found: C, 63.27; H, 5.92; N, 0.79.

2-(Trimethylsilyl)ethyl *O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-*O*-(6-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-*O*-benzoyl- β -D-glucopyranoside (11). To a solution of **8** (500 mg, 0.42 mmol) and **9** (420 mg, 0.72 mmol) in dry CH_3CN (5 mL) were added MS-3 \AA (3 g) and the mixture was stirred for 5 h at room temperature, and cooled to -40 $^\circ\text{C}$. NIS (562 mg, 2.5 mmol) and TfOH (64 μL , 0.72 mmol) were added, with stirring, to the mixture at -35 $^\circ\text{C}$ under N_2 atmosphere. A similar work-up as described for **10** gave **11** (405 mg, 58.2%) as an amorphous mass: $[\alpha]_{\text{D}} -8.8^\circ$ (c 1.8, CHCl_3); ^1H NMR (CDCl_3) δ 0.95 (m, 2H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.39 (d, 3H, $J_{5,6} = 6.2$ Hz, H-6b), 1.55, 1.82, 1.90, 2.00, 2.09 (5s, 15H, AcN, 4AcO), 2.54 (dd, 1H, $J_{\text{gem}} = 12.6$ Hz, $J_{3\text{eq},4} = 4.3$ Hz, H-3 deq), 4.29 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1c), 4.90 (m, 1H, H-4d), 5.10 (near t, 1H, $J_{2,3} = 10.0$ Hz, H-2a), 5.28 (dd, 1H, $J_{6,7} = 2.3$ Hz, $J_{7,8} = 9.5$ Hz, H-7d), 5.43 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1b), 5.56 (m, 1H, H-8d), and 7.30–8.23 (m, 30H, 6Ph).

Anal. Calcd for $\text{C}_{85}\text{H}_{101}\text{NO}_{30}\text{Si}$ (1644.8): C, 62.07; H, 6.19; N, 0.85. Found: C, 61.89; H, 5.91; N, 0.87.

2-(Trimethylsilyl)ethyl *O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-*O*-(2,4-di-*O*-acetyl-6-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-*O*-benzoyl- β -D-glucopyranoside (12). A solution of **11** (390 mg, 0.237 mmol) in EtOH (120 mL) and AcOH (20 mL) was hydrogenolyzed in the presence of 10% Pd-C (700 mg) for 2 days at 40 $^\circ\text{C}$, then filtered and concentrated. The residue was acetylated with Ac_2O (5 mL) in pyridine (10 mL) in the presence of 4-dimethylaminopyridine (10 mg) overnight at room temperature. The product was purified by column chromatography (100:1 CH_2Cl_2 -MeOH) on silica gel (50 g), to give **12** (340 mg, 89%) as an amorphous mass: $[\alpha]_{\text{D}} -6.0^\circ$ (c 1.9, CHCl_3); ^1H NMR (CDCl_3) δ 0.90 (m, 2H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.40 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.50, 1.74, 1.83, 1.99, 2.01, 2.03, 2.11 (2), 2.24, 2.29 (9s, 30H, AcN, 9AcO), 2.60 (dd, 1H, $J_{\text{gem}} = 12.3$ Hz, $J_{3\text{eq},4} = 4.2$ Hz, H-3 deq), 3.84 (s,

3H, MeO), 4.32 (d, 1H, $J_{1,2} = 8.2$ Hz, H-1c), 4.78 (d, 1H, $J_{1,2} = 7.2$ Hz, H-1a), 5.17 (dd, 1H, $J_{6,7} = 3.3$ Hz, $J_{7,8} = 7.2$ Hz, H-7d), 5.44 (d, 1H, $J_{1,2} = 3.8$ Hz, H-1b), 5.55 (d, 1H, $J_{3,4} = 3.0$ Hz, H-4c), and 7.22-8.28 (m, 15H, 3Ph).

Anal. Calcd for $C_{74}H_{107}NO_{35}Si$ (1598.7): C, 55.60; H, 6.75; N, 0.88. Found: C, 55.61; H, 6.86; N, 0.77.

***O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-*O*-(2,4-di-*O*-acetyl-6-*O*-benzoyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-acetyl- α -*L*-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-*O*-benzoyl-*D*-glucopyranose (13).** To a solution of **12** (191 mg, 0.12 mmol) in CH_2Cl_2 (2 mL) was added trifluoroacetic acid (2 mL) and the mixture was stirred for 3 h at 0 °C, then concentrated. Column chromatography (60:1 CH_2Cl_2 -MeOH) of the residue on silica gel (40 g) afforded **13** (171 mg, 95%) as an amorphous mass: $[\alpha]_D +10.0^\circ$ (*c* 1.7, $CHCl_3$); IR (KBr) 3380 (OH, NH), 1740 and 1230 (ester), 1640 and 1540 (amide), and 750 and 710 cm^{-1} (Ph); 1H NMR ($CDCl_3$) δ 1.34 (d, 1H, $J_{5,6} = 6.6$ Hz, H-6b), 1.42-2.35 (30H, AcN, 9AcO), 2.51 (dd, 1H, $J_{gem} = 12.6$ Hz, $J_{3eq,4} = 4.8$ Hz, H-3 deq), 3.75 (s, 3H, MeO), 4.90 (dt, 1H, H-4d), 5.17 (dd, 1H, $J_{6,7} = 2.2$ Hz, $J_{7,8} = 7.8$ Hz, H-7d), 5.26 (d, 1H, $J_{1,2} = 2.5$ Hz, H-1b), 5.47 (d, 1H, $J_{3,4} = 3.0$ Hz, H-4c), 5.57 (m, 1H, H-8d), and 7.12-8.18 (m, 15H, 3Ph).

Anal. Calcd for $C_{69}H_{95}NO_{35}$ (1498.5): C, 55.31; H, 6.39; N, 0.93. Found: C, 55.13; H, 6.65; N, 0.97.

***O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-*O*-(2,4-di-*O*-acetyl-6-*O*-benzoyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-acetyl- α -*L*-fucopyranosyl)-(1 \rightarrow 3)]-2,4-di-*O*-benzoyl- α -*D*-glucopyranosyl trichloroacetimidate (14).** A solution of **13** (70 mg, 47 μ mol) and trichloroacetonitrile (0.14 mL) in CH_2Cl_2 (1 mL) was cooled to 0 °C, and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 10 mg) was added. The mixture was stirred for 2 h at 0 °C and chromatographed on a column of silica gel (15 g) with 60:1 CH_2Cl_2 -MeOH, to give **14** (67 mg, 87%) as an amorphous mass: $[\alpha]_D +13.0^\circ$ (*c* 1.9, $CHCl_3$); 1H NMR ($CDCl_3$) δ 1.36 (d, 3H, $J_{5,6} = 6.6$ Hz, H-6b), 1.44, 1.68, 1.75, 1.93, 1.94 (2), 2.01 (2), 2.17, 2.21 (8s, 30H, AcN, 9AcO), 2.54 (dd, 1H, H-3 deq), 3.76 (s, 3H, MeO), 4.82 (m, 1H, H-4d), 5.45 (d, 1H, $J_{3,4} = 3.0$ Hz, H-4c), 5.47 (d, 1H, $J_{1,2} = 2.8$ Hz, H-1b), 6.35 (d, 1H, $J_{1,2} = 3.8$ Hz, H-1a), 7.15-8.17 (m, 15H, 3Ph), and 8.58 (s, 1H, C=NH).

Anal. Calcd for $C_{71}H_{97}N_2O_{35}Cl_3$ (1644.9): C, 51.84; H, 5.94; N, 1.73. Found: C, 51.75; H, 5.77; N, 1.53.

2-Azidoethyl *O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-*O*-(2,4-

di-*O*-acetyl-6-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-*O*-benzoyl- β -D-glucopyranoside (15). To a solution of **14** (67 mg, 41 μ mol) and 2-azidoethanol (8.6 mg, 90 μ mol) in dry CH_2Cl_2 (0.5 mL) were added MS-4 \AA (AW-300, 200 mg) and the mixture was stirred for 5 h at room temperature, then cooled to 0 $^\circ\text{C}$. Boron trifluoride etherate (3 μ L) was added and the mixture was stirred for 8 h at 0 $^\circ\text{C}$ and then filtered. Dichloromethane (50 mL) was added, and the solution was successively washed with M Na_2CO_3 and H_2O , dried (Na_2SO_4), then concentrated. Column chromatography (60:1 CH_2Cl_2 -MeOH) of the residue on silica gel (20 g) gave **15** (58 mg, 91%) as an amorphous mass: $[\alpha]_{\text{D}} -4.5^\circ$ (*c* 1.2, CHCl_3); IR (KBr) 3350 (NH), 2100 (N_3), 1740 and 1230 (ester), 1670 and 1540 (amide), and 750 and 710 cm^{-1} (Ph); ^1H NMR (CDCl_3) δ 1.33 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.42, 1.66, 1.75, 1.92, 1.94, 1.96, 2.03, 2.06, 2.17, 2.22 (10s, 30H, AcN, 9AcO), 3.77 (s, 3H, MeO), 4.32 (d, 1H, $J_{1,2} = 8.0$ Hz, H-1c), 4.79 (d, 1H, $J_{1,2} = 7.5$ Hz, H-1a), 5.35 (d, 1H, $J_{1,2} = 3.0$ Hz, H-1b), 5.47 (d, 1H, $J_{3,4} = 2.8$ Hz, H-4c), and 7.27-8.20 (m, 15H, 3Ph).

Anal. Calcd for $\text{C}_{71}\text{H}_{98}\text{N}_4\text{O}_{35}$ (1567.6): C, 54.40; H, 6.30, N, 3.57. Found: C, 54.66; H, 6.32; N, 3.65.

8-Azidoethyl *O*-(Methyl 5-Acetamido-4, 7, 8, 9-tetra-*O*-acetyl-3, 5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-*O*-(2,4-di-*O*-acetyl-6-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-*O*-benzoyl- β -D-glucopyranoside (16). Coupling of **14** (52 mg, 32 μ mol) and 8-azidoethanol (8 mg, 47 μ mol), as described for **15**, gave **16** (43 mg, 78%) as an amorphous mass: $[\alpha]_{\text{D}} -8.0^\circ$ (*c* 0.8, CHCl_3); IR (KBr) 3350 (NH), 2940 (CH_2), 2100 (N_3), 1750 and 1230 (ester), 1660 and 1540 (amide), and 710 cm^{-1} (Ph); ^1H NMR (CDCl_3) δ 1.34 (d, 3H, $J_{5,6} = 6.6$ Hz, H-6b), 1.43-2.20 (10s, 30H, AcN, 9AcO), 2.52 (dd, 1H, $J_{\text{gem}} = 12.8$ Hz, $J_{3\text{eq}, 4} = 4.9$ Hz, H-3 deq), 3.76 (s, 3H, MeO), 4.37 (d, 1H, $J_{1,2} = 8.3$ Hz, H-1c), 4.84 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1a), 5.46 (d, 1H, $J_{1,2} = 2.6$ Hz, H-1b), and 7.18-8.23 (m, 15H, 3Ph).

Anal. Calcd for $\text{C}_{77}\text{H}_{110}\text{N}_4\text{O}_{35}$ (1651.7): C, 55.99; H, 6.71, N, 3.39. Found: C, 56.14; H, 6.75; N, 3.38.

2-[2-(2-Azidoethoxy)ethoxy]ethyl *O*-(Methyl 5-Acetamido-4,7,8,9-tetra-*O*-acetyl-3, 5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-*O*-(2,4-di-*O*-acetyl-6-*O*-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(2,3,4-tri-*O*-acetyl- α -L-fucopyranosyl)-(1 \rightarrow 3)]-2,6-di-*O*-benzoyl- β -D-glucopyranoside (17). Coupling of **14** (95.5mg, 58 μ mol) and 2-[2-(2-azidoethoxy)ethoxy]ethanol (15 mg, 87 μ mol), as described for **15**, gave **17** (76 mg,

79%) as an amorphous mass: $[\alpha]_D -4.3^\circ$ (*c* 1.5, CHCl_3); IR (KBr) 3350 (NH), 2120 (N_3), 1750 and 1220 (ester), 1660 and 1540 (amide), and 760 and 720 cm^{-1} (Ph); ^1H NMR (CDCl_3) δ 1.32 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.34, 1.42, 1.64, 1.75, 1.92, 1.94, 1.95, 2.04, 2.16, 2.21 (10s, 30H, AcN, 9AcO), 2.52 (dd, 1H, $J_{\text{gem}} = 12.8$ Hz, $J_{3eq,4} = 4.9$ Hz, H-3deq), 3.76 (s, 3H, MeO), 4.33 (d, 1H, $J_{1,2} = 7.5$ Hz, H-1c), 4.78 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1a), 5.35 (dd, 1H, $J_{6,7} = 2.0$ Hz, $J_{7,8} = 9.5$ Hz, H-7d), 5.47 (d, 1H, $J_{1,2} = 2.5$ Hz, H-1b), and 7.28-8.20 (m, 15H, 3Ph).

Anal. Calcd for $\text{C}_{75}\text{H}_{106}\text{N}_4\text{O}_{37}$ (1655.7): C, 54.41; H, 6.45, N, 3.39. Found: C, 54.43; H, 6.39; N, 3.33.

2-Azidoethyl *O*-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-*O*-(β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(α -L-fucopyranosyl)-(1 \rightarrow 3)]- β -D-glucopyranoside (18). To a solution of **15** (58 mg, 37 μmol) in MeOH (5 mL) was added NaOMe (20 mg) and the mixture was stirred overnight at 40 $^\circ\text{C}$; the course of the reaction was monitored by TLC. Water (3 mL) was added to the mixture, and this was stirred overnight at room temperature, neutralized with Amberlite IR-120 (H^+) resin and filtered. The resin was washed with MeOH, and the combined filtrate and washings were concentrated. Column chromatography (MeOH) of the residue on Sephadex LH-20 (40 g) gave **18** (33 mg, quantitative) as an amorphous mass: $[\alpha]_D -39.0^\circ$ (*c* 0.9, MeOH); ^1H NMR (CD_3OD) δ 1.27 (d, 3H, $J_{5,6} = 6.6$ Hz, H-6b), 2.11 (s, 3H, AcN), 2.55 (broad dd, 1H, H-3deq), 4.44 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1c), 4.58 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1a), and 5.56 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1b).

Anal. Calcd for $\text{C}_{31}\text{H}_{52}\text{N}_4\text{O}_{23}$ (848.8): C, 43.87; H, 6.18, N, 6.60. Found: C, 44.06; H, 6.35; N, 6.54.

8-Azidoethyl *O*-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-*O*-(β -D-galactopyranosyl)-(1 \rightarrow 4)-[*O*-(α -L-fucopyranosyl)-(1 \rightarrow 3)]- β -D-glucopyranoside (19). Deacylation and saponification of **16** (42.5 mg, 24.7 μmol), as described for **18**, yielded compound **19** (25 mg, quantitative) as an amorphous mass: $[\alpha]_D -33.7^\circ$ (*c* 0.9, MeOH); ^1H NMR (CD_3OD) δ 1.07 (d, 3H, $J_{5,6} = 6.6$ Hz, H-6b), 1.18-1.69 (m, 12H, 6 CH_2), 1.91 (s, 3H, AcN), 2.37, 3.18 (m, 4H, CH_2), 2.78 (broad dd, 1H, H-3deq), 4.17 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1c), 4.38 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1a), and 5.36 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1b).

Anal. Calcd for $\text{C}_{37}\text{H}_{64}\text{N}_4\text{O}_{23}$ (932.9): C, 47.64; H, 6.92, N, 6.01. Found: C, 47.52; H, 7.12; N, 5.81.

2-[2-(2-Azidoethoxy)ethoxy]ethyl *O*-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)-*O*-(β -D-galac-

topyranosyl)-(1→4)-[O-(α-L-fucopyranosyl)-(1→3)]-β-D-glucopyranoside (**20**). Deacylation and saponification of **17** (76 mg, 46 μmol), as described for **18**, afforded compound **20** (45 mg, quantitative) as an amorphous mass: $[\alpha]_D -34.0^\circ$ (*c* 1.5, MeOH); $^1\text{H NMR}$ (CD₃OD) δ 1.07 (d, 3H, $J_{5,6} = 6.4$ Hz, H-6b), 1.92 (s, 3H, AcN), 2.78 (broad dd, 1H, H-3deq), 3.40-3.90 (m, 12H, 6CH₂), 4.24 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1c), 4.38 (d, 1H, $J_{1,2} = 7.7$ Hz, H-1a), and 5.37 (d, 1H, $J_{1,2} = 3.5$ Hz, H-1b).

Anal. Calcd for C₃₅H₆₀N₄O₂₅(936.9): C, 44.87; H, 6.46, N, 5.98. Found: C, 44.69; H, 6.71; N, 5.83.

2-(Trimethylsilyl)ethyl O-(5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyranosylonic acid)-(2 → 3)-O-(β-D-galactopyranosyl)-(1→4)-[O-(α-L-fucopyranosyl)-(1→3)]-β-D-glucopyranoside (**21**). Deacylation and saponification of **12** (71.5 mg, 44.4 μmol), as described for **18**, gave compound **21** (39 mg, quantitative) as an amorphous mass: $[\alpha]_D -19.1^\circ$ (*c* 1.3, MeOH); $^1\text{H NMR}$ (CD₃OD) δ 0.98 (m, 2H, MeSiCH₂CH₂), 1.13 (d, 3H, $J_{5,6} = 6.2$ Hz, H-6b), 1.99 (s, 3H, AcN), 2.83 (broad dd, 1H, H-3deq), 4.26 (d, 1H, $J_{1,2} = 7.9$ Hz, H-1c), 4.45 (d, 1H, $J_{1,2} = 8.4$ Hz, H-1a), and 5.45 (d, 1H, $J_{1,2} = 3.7$ Hz, H-1b).

Anal. Calcd for C₃₄H₆₁NO₂₃Si(879.9): C, 47.64; H, 6.99, N, 1.59. Found: C, 47.52; H, 7.15; N, 1.47.

ACKNOWLEDGMENT

This work was supported in part by a Grant-in-Aid (No. 04250102 and 03660132) for Scientific Research on Priority Areas from the Ministry of Education, Science and Culture of Japan.

REFERENCES

1. M. L. Phillips, E. Nudelman, F. C. A. Gaeta, M. Perez, A. K. Singhal, S. Hakomori and J. C. Paulson, *Science*, **250**, 1130 (1990).
2. G. Walz, A. Aruffo, W. Kolanus, M. Bevilacqua and B. Seed, *Science*, **250**, 1132 (1992).
3. D. Tyrrell, P. James, N. Rao, C. Foxall, S. Abbas, F. Dasgupta, M. Nashed, A. Hasegawa, M. Kiso, D. Asa, J. Kidd and B. K. Brandley, *Proc. Natl. Acad. Sci. USA*, **88**, 10372 (1991).
4. a) A. Kameyama, H. Ishida, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, **209**, c1 (1991); b) *ibid.*, *J. Carbohydr. Chem.*, **10**, 549 (1991).

5. a) A. Hasegawa, T. Ando, A. Kameyama and M. Kiso, *Carbohydr. Res.*, **230**, c1 (1992); b) *ibid.*, *J. Carbohydr. Chem.*, **11**, 645 (1992).
6. A. Kameyama, H. Ishida, M. Kiso and A. Hasegawa, *J. Carbohydr. Chem.*, **10**, 729 (1991).
7. a) H. Furui, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, **229**, c1 (1992) ; b) M. Yoshida, A. Uchimura, M Kiso, and A. Hasegawa, *Glycoconjugate J.*, in press (1993); c) M. Yoshida, T. Ando, A. Uchimura, M. Kato, M. Kiso and A. Hasegawa, Presented at the *XVIth International Carbohydrate Symposium*, July 5-10, 1992, Paris, France, ABSTRACTS BOOK p 112.
8. M. Larkin, T. J. Ahern, M. S. Stoll, M. Shaffer, D. Sako, J. O'Brien, C.-T. Yuen, A. M. Lawson, R. A. Childs, K. M. Barone, P. R. Langer-Safer, A. Hasegawa, M. Kiso, G. R. Larsen and T. Feizi, *J. Biol. Chem.*, **267**, 13661 (1992).
9. C. Foxall, S. R. Matson, D. Dowbenko, C. Fennie, L. A. Lasky, M. Kiso, A. Hasegawa, D. Asa and B. K. Brandley, *J. Cell Biol.*, **117**, 895 (1992).
10. A. Tanaka, K. Ohmori, N. Takahashi, K. Tsuyuoka, A. Yago, K. Zenita, A. Hasegawa and R. Kannagi, *Biochem. Biophys. Res. Commun.*, **179**, 713 (1991).
11. K. Ohmori, A. Tanaka, T. Yoneda, Y. Buma, K. Hirashima, K. Tsuyuoka, A. Hasegawa and R. Kannagi, *Blood*, **81**, 101 (1993).
12. B. K. Brandley, M. Nashed, F. Dasgupta, S. Abbas, D. Asa, C. Foxall and J. Musser, Presented at *Keyston Symposia*, Jan. 24-31, 1993, Keystone, CO, USA.
13. a) K. Jansson, S. Ahlfors, T. Frejd, J. Kihlberg, G. Magnusson, J. Dahmen, G. Noori and K. Stenvall, *J. Org. Chem.*, **53**, 5629 (1988); b) K. P. R. Kartha, A. Kameyama, M. Kiso and A. Hasegawa, *J. Carbohydr. Chem.*, **8**, 145 (1989).
14. P. Fügedi and P. J. Garegg, *Carbohydr. Res.*, **149**, c9 (1986).
15. F. Yamazaki, S. Sato, T. Numata, Y. Ito and T. Ogawa, *Carbohydr. Res.*, **201**, 31 (1990).
16. A. Marra and S. Sinay, *Carbohydr. Res.*, **187**, 35 (1989).
17. a) G. H. Veeneman, S. H. van Leeuwen and J. H. van Boom, *Tetrahedron Lett.*, **31**, 1131 (1990); b) P. Konradsson, D. R. Mootoo, R. E. McDevitt and B. Fraser-Reid, *J. Chem. Soc. Chem. Commun.*, 270 (1990); c) A. Hasegawa, T. Nagahama, H. Ohki, K. Hotta, H. Ishida and M. Kiso, *J. Carbohydr. Chem.*, **10**, 493 (1991).
18. a) T. Murase, H. Ishida, M. Kiso and A. Hasegawa, *Carbohydr. Res.*, **184**, c1 (1988); b) *ibid.*, **188**, 71 (1989); c) T. Murase, A. Kameyama, K. P. R. Kartha, H. Ishida, M. Kiso and A. Hasegawa, *J. Carbohydr. Chem.*, **8**, 265 (1989); d) A. Hasegawa, H. Ohki, T. Nagahama, H. Ishida and M. Kiso, *Carbohydr. Res.*, **212**, 277 (1991).
19. a) R. R. Schmidt and G. Grundler, *Synthesis*, 885 (1981); b) Y. Ito, M. Kiso and A. Hasegawa, *J. Carbohydr. Chem.*, **8**, 285 (1989).